



## Thaw and Culture Details

Cell Line Name	<b>JHU102i</b>
WiCell Lot Number	<b>DB41279</b>
Provider	Johns Hopkins University – Laboratory of Dr. Lewis Becker
Banked By	Johns Hopkins University – Laboratory of Dr. Lewis Becker
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 4 wells of a 6 well plate using TeSR™-E8™ and Recombinant Human Vitronectin. WiCell recommends thawing using ROCK Inhibitor for best results.
Protocol	WiCell Feeder Independent Pluripotent Stem Cell Protocol
Culture Platform Prior to Freeze	Feeder Independent
	Medium: E8
	Matrix: Vitronectin
Passage Number	p4 These cells were cultured for 4 passages post reprogramming prior to freeze. Add +1 to the passage number to best represent the overall passage number of the cells at thaw.
Date Vialied	04-June-2016
Vial Label	P102 P4 6/4/16 0.9M
Biosafety and Use Information	This cell line is of human origin. Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

## Testing Performed by WiCell

Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype by G-banding	WiCell	SOP-49	Expected karyotype	See Report
Post-Thaw Viable Cell Recovery	WiCell	SOP-99	Recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega	Defines STR profile of deposited cell line	Pass
Sterility	Steris	ST/07	Negative	Pass
Mycoplasma	WiCell	SOP-79	Negative	Pass

## Testing Reported by Provider

The Provider stated that some or all of the additional analyses listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

- Embryoid bodies
- Infinium® Expanded Multi-Ethnic Genotyping Array (MEGA<sup>EX</sup>)



Approval Date	Quality Assurance Approval
26-August-2016	<p style="text-align: right;">12/3/2020</p> <p>X JKG</p> <hr/> <p><small>JKG Quality Assurance Signed by: Gay, Jenna</small></p>

**Date Reported:** Friday, November 13, 2020

**Cell Line:** JHU102i-DB41279

**Submitted Passage #:** 5

**Date of Sample:** 11/6/2020

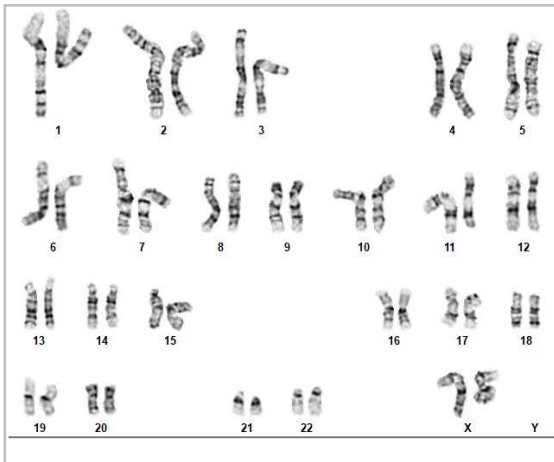
**Specimen:** Human iPSC

**Results:** 46,XX

**Cell Line Sex:** Female

**Reason for Testing:** LOT\_RELEASE

**Investigator:** WiCell Stem Cell Bank, WiCell



**Cell:** 35

**Slide:** G01

**Slide Type:** Karyotype

**Total Counted:** 20

**Total Analyzed:** 8

**Total Karyogrammed:** 4

**Band Resolution:** 400 - 450

**Interpretation:**

**This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.**

**Completed by:** [REDACTED], CG(ASCP)

**Reviewed and Interpreted by:** [REDACTED] Ph.D.

**Date:** \_\_\_\_\_ **Sent By:** \_\_\_\_\_ **Sent To:** \_\_\_\_\_ **QC Review By:** \_\_\_\_\_

*Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".*

*This assay was conducted solely for listed investigator/institution. The results of this assay are for research use only. Unless otherwise mutually agreed in writing, the services provided to you hereunder by WiCell Research Institute, Inc. ("WiCell") are governed solely by WiCell's Terms and Conditions of Service, found at [www.wicell.org/privacyandterms](http://www.wicell.org/privacyandterms). Any terms you may attach to a purchase order or other document that are inconsistent, add to, or conflict with WiCell's Terms and Conditions of Service are null and void and of no legal force or effect.*



# Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell  
 Samples Received: 02Nov20, 06Nov20  
 STR Amplification Date: 11Nov20

Sample Name	WA09-RB67589 p.30	JHU105i- DB36241 p.5	JHU148i-DB36280 p.7	JHU102i-DB41279 p.5	JHU250i- DB36904 p.8	JHU173i-DB36380 p.10
Label on tube	83593	83677	83678	83679	83687	83688
FGA	Identifying information has been redacted to protect donor confidentiality. If more information is required, please contact <a href="mailto:info@wicell.org">info@wicell.org</a>					
TPOX						
D8S1179						
vWA						
Amelogenin						
Penta_D						
CSF1PO						
D16S539						
D7S820						
D13S317						
D5S818						
Penta_E						
D18S51						
D21S11						
TH01						
D3S1358						
Allelic Polymorphisms	24	30	27	25	25	43
Matches*	See Matches Comment					
Comments						See Mixed Cell Line Comment

*\*Note: The STR profile of the following sample is an exact match for the given sample/samples.*



# Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell  
Samples Received: 02Nov20, 06Nov20  
STR Amplification Date: 11Nov20

**Results:** The genotypic profiles comprise a range of 24-43 allelic polymorphisms across the 15 STR loci analyzed.

**Interpretation:** The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggests that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

**Sensitivity:** Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-5%.

**Mixed Cell Line:** Sample 83688 shows signs of possible contamination. The most likely explanation for this result is that two cultures have been mixed. G-banded karyotype results confirmed the presence of both male and female cells in the culture. Please resubmit this sample.

**Matches:** Sample 83593 is an exact match to 14630, 74319, 74844, 74924, and 74925.

12/1/2020

12/1/2020

12/1/2020

X [Redacted]

Tech #1

Characterization

Signed by: [Redacted]

X [Redacted]

Tech #2

Characterization

Signed by: [Redacted]

X [Redacted]

QA Review

Quality Assurance

Signed by: [Redacted]

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*Raw data is available upon request.*

# Native Product Sterility Report



WiCell  
504 S Rosa Road, Rm 101  
Madison, WI 53719

SAMPLE #: 18121076  
DATE RECEIVED: 13-Dec-18  
TEST INITIATED: 17-Dec-18  
TEST COMPLETED: 31-Dec-18

SAMPLE NAME / DESCRIPTION: LUEL8363i-3 WB66952 14169, JHU088i DB41240 14170, JHU132i DB36272 14171, JHU102i DB41279 14172, JHU103i DB36236 14173, iPS DF19-9-7T DB36826 14174, iPS DF4-3-7T.A DB36837 14175, iPS DF6-9-9T.B DB36844 14176, MIRJT7i-mND2-0 DB36854 14177, IISH1i-BM1 DB36864 14178, IISH2i-BM9 DB36871 14179, IISH3i-CB6 DB36881 14180, WA01 DB36785 14181, WA07 DB36888 14182, WA09 DB36899 14183, WA24 DB36798 14184, JHU155i DB41365 14185, JHU184i DB41392 14186, JHU142i DB41344 14187, WA09 NA65899 14189.

UNIQUE IDENTIFIER: NA

## TEST RESULTS:

# Tested	# Positives (Growth)	- Control
20	1	2 Negatives

## TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
20	TSB	40	20-25	14
20	FTG	40	30-35	14

REFERENCE: Processed according to LAB-003: Sterility Test Procedure

PD #: 000053

TEST METHODOLOGY: USP - Direct Transfer

COMMENTS: Sample labeled P142 P6 6/16/16 0.9M is positive in TSB and FTG media.

Reported as per packing slip.

REVIEWED BY \_\_\_\_\_

DATE 02 JAN 19

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests. Results applied to samples as received.



# Mycoplasma Assay Report

PCR-based assay performed by WiCell

WiCell

04Nov20

FORM SOP-83.01

Version 01

Sample Name	Result	Comments/Suggestions
INC149 02Nov20AP (83598)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC123 02Nov29KR 1 of 2 (83599)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC123 02Nov20KR 2 of 2 (83600)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC169 02Nov20MMM 1 of 2 (83601)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC169 02Nov20MMM 2 of 2 (83602)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU105i-DB36241 (83622)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU004i-2-DB40945 (83623)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU036i-DB40981 (83624)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU102i-DB41279 (83625)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU148i-DB36280 (83626)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU173i-DB36380 (83627)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU214i-DB36851 (83628)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU234i-DB37041 (83629)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU250i-DB36904 (83630)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

Reported by: [REDACTED], Senior Cell Culture Specialist

Reviewed by: [REDACTED] Assistant Cell Culture Specialist

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*A gel image is available upon request.*